
ASSESSMENT OF GENETIC DISTANCE AND SIMILARITIES AMONG INDIGENOUS CHICKEN POPULATIONS IN KANO-STATE, NIGERIA

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ABSTRACT

The study was conducted to examine the genetic diversity of three populations of indigenous chicken in Kano State, Nigeria. A total of 249 matured free-ranging chickens between the ages of 6 -24 months were sampled. Three genetic groups of indigenous chicken were identified in the study location which includes normal feathered (Nf), frizzle feathered (Ff) and naked neck (Na) and 2mL of blood samples obtained from 8 birds per genotype were used for the study. Genomic DNA was extracted individually from the sampled blood using Quick DNA Kits (©Zymo-Spin Research Kit). Five microsatellite markers were used for genotyping. Genetics Analysis in Excel Software (GenA1EX software version 6.5) was used for band analysis. The result showed relatively high genetic diversity which is essential for the populations. The chicken populations had degrees of genetic similarity of (0.840-0.925) and genetic distance of (0.078-0.084). High genetic distance of (0.084) was observed between Nf and Ff. It was concluded that the populations were not isolated and the microsatellite markers used were highly informative and can be used in future studies involving chicken populations.

Keyword: Genetic Distance, Genetic Similarity, Indigenous Chickens

INTRODUCTION

Domestic chicken (*Gallus gallus domesticus*) was estimated to have chromosome number as $2n = 78$ (Pollock and Fechheimer, 1976), showing the sex chromosomes Z and W, and female as heterogametic sex (Nakamura *et al.*, 2004). Indigenous chickens (IC) express various genes for egg and meat production, heat tolerance, feed efficiency, disease resistance, growth rate and feather type (Singh *et al.*, 2001). The existence of various ecotypes was due to the product of mutation and recombination, genetic drift, adaptation, migration, selection pressures, artificial selection, activities of man and available nutrition (Barker, 1994). IC have been reported to exhibit high genetic variability which indicate their potential for genetic improvement (Dana and Vander Waaij, 2011). The Nigerian indigenous chicken is a dual-purpose bird that is used both for meat and egg production in the rural and peri-urban areas of the country. They are found in large numbers distributed across different agroecological categories under a traditional family based scavenging management system (Sonaiya and Olori, 1990). The IC are well adapted to the adverse tropical conditions, stresses diseases resistance and low management inputs (Ikeobi *et al.*, 1996; Adeleke *et al.*, 2011). The IC contain a highly conserved genetic system with high levels of heterozygosity (Wimmers *et al.*, 2000). Genetic diversity is the variation of alleles and genotype present in a population (Sheriff and Alemayehu, 2017) and it is the product of interaction between environment and genetic effects which leads to differentiation of morphological, physiological and productive traits that provide selection criterion for genetic improvement (Gambo *et al.*, 2022). The objective of this study was to evaluate an existing genetic variation within and among the IC populations in Kano State. Kano is one of the oldest cities in Nigeria. It is located at the Northern region specifically at North-western part of Nigeria. It has a total land area of 20,131km² (Khalil *et al.*, 2015).

MATERIALS AND METHODS

The study was conducted at eight randomly selected locations in Kano State, Nigeria. They include; Minjibir, Kumbotso, Gezawa, Dawakin kudu, Wudil, Kura, Madobi and Tudun wada. Kano State is a typical tropical area located between latitudes 10°30'N and 12°38'N and longitudes 7°45'E and 9°29'E. The three genetic groups: Na, Ff and Nf were phenotypically identified in the selected villages. Twenty-four (24) blood samples (eight samples from each genotype) were randomly selected for the analysis. Two millilitres (2mL) of blood was collected from brachial vein using a 2mL sterile syringe and stored in Ethylene Di-amine Tetra Acetic Acid (EDTA) tube. The birds were physically restrained to avoid injury. All experimental procedures were carried out in accordance with the guidelines of the

Institutional Animal Ethics Committee (IAEC). Laboratory analysis for DNA extraction, purification and amplification were done at the National Veterinary Research Institute (N.V.R.I), Jos, Plateau State.

DNA Extraction and Polymerase Chain Reaction (PCR)

Genomic DNA was extracted individually from the sampled blood using Zymo-Spin research kit (ZR kit/DNA MiniPrep) according to the manufacturer's procedure. Five microsatellite loci which are among the suggested markers by FAO (2011) were used for genotyping. The primers were ADL0268, LEI0166, MCW0183, MCW0222 and MCW0067.

Electrophoresis

Gel electrophoresis was conducted on 1% Agarose gel to separate DNA according to size. DNA is negatively charged and is drawn towards the positive electrode. Smaller fragments moved down the gel faster and moved through the gel matrix. Thus, allows band to be distinguished which differs in its size. PCR products were genotyped to determine specific allele.

Statistical Analysis

Nei's genetic distance and identity per population was computed, the bands were subjected to GenAlex software version 6.5 (Peakall and Smouse, 2012). The population matrix was used to compared the distance between the different population and indicate their relationship. The result was compared with Nei's (1978) unbiased genetic distance and identity calculated to assess their genetic diversity.

RESULTS AND DISCUSSION

Genetic Similarities

The similarity values for the IC populations studied were presented in table 1. The table showed that *Nf* was related to *Na* with a similarity value of 0.925. It was observed that the similarity value between *Nf* and *Ff* is 0.919. Lowest similarity value was found between *Na* and *Ff* (0.840). Table 2 represented the unbiased genetic similarity values when all sources of error are removed from analysis. Adeleke *et al.*, (2011) observed genetic similarities of 0.4983, 0.3632 and 0.441 between *Na* and *Ff*, *Nf* and *Ff*, *Na* and *Nf* respectively. However, high genetic similarity estimates maybe because of the effect of small population size, high selection pressure in closed population, inbreeding and minimal or null immigration of new genetic materials into the population (Canon *et al.*, 2006). According to Abebe, Mikko and Johansson (2015), mating individuals that are genetically identical can also result to a high genetic similarity.

Genetic Distances

The genetic distance of the chicken populations studied was presented in table 3. Highest value was observed between *Na* and *Ff* (0.174). The unbiased genetic distance values give the when all sources of error are removed from the analysis represented in table 4. The result of the study is lower than report of Adeleke *et al.*; (2011) who found the genetic distance of (0.5585) between *Nf* and *Na* and the distance between *Nf* and *Ff* is 0.6368. Wimmers *et al.*; (2000) observed genetic distance between the Nigerian chickens ranged from 0.05 to 0.15. Okamoto *et al.*; (1999) reported small genetic differentiation among the Laos chicken populations studied. Hillel *et al.*; (2003) showed that genetic distance measures based on gene frequencies were in agreement with the genetic diversity of these populations, indicating that these approaches fit the history of domesticated chickens well.

Table 1: Nei's Genetic Similarities among Population of Indigenous Chicken Populations

	Nf	Na	Ff
Nf	1.000		
Na	0.925	1.000	
Ff	0.919	0.840	1.000

Table 2: Nei's Unbiased Genetic Similarities among Population of Indigenous Chicken

	Nf	Na	Ff
Nf	1.000		
Na	0.984	1.000	
Ff	0.974	0.888	1.000

Table 3: Nei's Genetic Distance among Population of Indigenous Chicken

	Nf	Na	Ff
Nf	0.000		
Na	0.078	0.000	
Ff	0.084	0.174	0.000

Table 4: Nei's Unbiased Genetic Distance among Population of Indigenous Chicken

	NF	Na	Ff
NF	0.000		
Na	0.016	0.000	
Ff	0.026	0.119	0.000

CONCLUSION

The study showed relatively high genetic diversity, thus are good reservoir of allelic diversity which is essential for the populations to be more adaptive to environmental changes. The chicken populations had degrees of genetic similarity of (0.840-0.925) and genetic distance of (0.078-0.084) and none of the genotype exhibited genetic uniqueness. Therefore, it can be concluded that the populations were not isolated and can be used for genetic improvement.

REFERENCES

- Abebe, A.S., Mikko S., Johansson, A.M. (2015).** Genetic Diversity of Five Local Swedish Chicken Breeds Detected by Microsatellite Markers. *Public Library of Science One*: 10(4): E0120580.
- Adeleke, M.A., Peters, S.O., Ozoje, M.O., Ikeobi, C.O.N., Adebambo, A.O., Olowofeso, O., Bamgbose, A.M. and Adebambo, O.A. (2011).** A Preliminary Screening of Genetic Lineage of Nigerian Indigenous Chickens Based on Blood Protein Polymorphisms. *Animal Genetic Resources*, 48:23–28.
- Barker, J.S.F. (1994).** A Global Protocol for Determining Genetic Distances among Domestic Livestock Breeds. *Proceedings of 5th World Congress on Genetics Applied to Livestock Production*, 21; 501-508.
- Canon, J., Garcí'a, D., Garcí'a-Atance, M.A., Obexer-Ruff, G., Lenstra, J.A., Ajmone-Marsan, P. and Dunner, S. (2006).** The ECONOGENE Consortium. Geographical partitioning of goat diversity in Europe and the Middle East. *Journal of Animal Genetics*, 37: 327334.
- Dana N. and Vander Waaij, E. H. (2011).** Genetic and Phenotypic Parameter Estimates for Bodyweight and Egg Production in Horro Chicken of Ethiopia. *Journal of Tropical Animal Health and Production*, 43(1): 21-28.
- FAO. (2011).** Molecular Genetic Characterization of Animal Genetic Resources. FAO Animal Production and Health Guidelines. No.9. Rome, Italy.
- Gambo, D, Faith, E.A. and Abdullahi, J. (2022).** Testing of Hardy-Weinberg Equilibrium in Population of Two Nigerian Local Chicken Ecotypes Base on Location. *EAS Journal of Biotechnology and Genetics*, 4(6): 80-83.
- Hillel, J., Groenen, M.A.M., Tixier-Boichard, M., Korol, A.B., David, L., Kirzhner, V.M., Burke, T., Barre-Dirie, A. (2003).** Biodiversity of 52 Chicken Populations Assessed by Microsatellite Typing of DNA Pools. *Genetics Selection Evolution*, 35: 533-557.
- Khalil, M.S., Sabiu, N., Muhammad, S.N (2015).** A Brief Look at Geography Education in Nigeria: A Case Study of Kano State. *Dutse Journal of Pure and Applied Sciences* 1(1) :11-18.
- Ikeobi, C.O.N., Ozoje, M.O., Adebambo, O.A.J., Adenowo, A. and Adebambo, O.A. (1996).** Genetic differences in the performance of local chicken in south-western Nigeria. *Nigerian Journal of Genetics*, 11: 32-39.
- Nakamura, D., Tiersch, T.R., Douglass, M. and Chandler, R.W. (2004).** Rapid Identification of Sex in Birds by Flow Cytometry, *Cytogenetics Cell Genetics*, 53(4):201-205.
- Nei, M. (1978).** Estimation of average heterozygosity and genetic distance from a small number of individuals. *Journal of Genetics*, 89: 583-590.
- Okamoto, S., Tsunekawa, N., Kawamoto, Y., Worawut, R., Kawabe, K., Maeda, Y. and Nishida, T. (1999).** Blood Protein Polymorphisms of Native Fowl in Laos. *Asian-Australas Journal of Animal Science*, 12; 1011-1014.

- Peakall, R. and Smouse, P.E. (2012).** GenAlEx 6.5: Genetic Analysis in Excel. Population Genetic Software for Teaching and Research – an update. *Bioinformatics*, 28(19): 2537-2539.
- Pollock, D.L. and Fechheimer N.S. (1976).** The Chromosome Number of *Gallus domesticus*. *Journal of Poultry Science*, 17(1):39–42.
- Sheriff, O. and Alemayehu, K. (2017).** Genetic Diversity Studies Using Microsatellite Markers and Their Contribution in Supporting Sustainable Sheep Breeding Programs. *Asian Journal of Agriculture*, 1(1): 46-51.
- Singh, C.V., Kumar, D. And Singh Y.P. (2001).** Potential Usefulness of the Plumage Reducing Naked Neck (Na) Gene in Poultry Production at Normal and High Ambient Temperature. *World Poultry Science Journal*, 57: 139-156.
- Sonaiya, E.B and Olori, V.E. (1990).** Family Poultry Production in South-western Nigeria. Proceedings of International workshop (pp. 243-247). Ile Ife Nigeria: Obafemi Awolowo University.
- Wimmers, K., Siriluck, P., Friedrich, S., Torsten, H.E., Babafunso, S., Karl, S., Peter H. (2000).** Application of Microsatellite Analysis to Group Chicken According to their Genetic Similarity. *Arch. Tierz., Dummerstorf*, 42(6): 629-639